

IRSTI 578.5

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SYNTHESIS OF PRIMERS AND DEVELOPMENT OF SIGNIFICANT GENES OF B.1.1.7 (ALPHA) VARIANT OF THE SARS-COV-2 VIRUS

Abstract. The threat of human infection with coronavirus became known back in the 60s of the last century. Since then, zoonotic coronavirus infection has spread among people and there were two major epidemics at the beginning of the 21st century. The rate of human infection with coronavirus infection spread rapidly and caused serious concern in the world. Coronavirus infection has undergone various genetic changes over time, as a result, in 2019, in Wuhan, China, coronavirus infection spread among people. Due to the widespread spread of coronavirus infection in the world, *WHO* declared a pandemic on March 11, 2020. Coronavirus infection *SARS-CoV-2* has been registered in about 200 countries around the world and has undergone various genetic changes over time. Due to genetic changes, a new strain of coronavirus infection appeared in the *UK* in the autumn of 2020. To determine the genetic changes of coronavirus infection and to study them, 65 pairs of primers were developed in the Collective Use Laboratory to obtain the complete nucleotide sequence of the *SARS-CoV-2* virus. Specific primers have been developed for the development of significant genes of the *SARS-CoV-2* virus. The developed primers allow us to obtain complete information about the genes of *SARS-CoV-2* viruses.

The presented primers can be used for identification and development of the complete genome of the *SARS-CoV-2* virus

Keywords: *SARS-CoV-2*, mutation, PCR.

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ПРАЙМЕРЛЕР СИНТЕЗІ ЖӘНЕ SARS-COV-2 ВИРУСЫ B.1.1.7 (АЛЬФА) НҰСҚАСЫНЫҢ МАҢЫЗДЫ ГЕНДЕРДІН ӨНДЕУ

Аннотация. Адамның коронавирус жұқтыру қауіпі өткен ғасырдың 60-жылдарында белгілі болды. Содан бері зоонозды коронавирустық инфекция адамдарға таралып, 21 ғасырдың басында екі үлкен індет болды. Адамдардың коронавирустық инфекцияны жұқтыру көрсеткіші тез таралды және әлемде үлкен алаңдаушылық тудырды. Коронавирустық инфекция уақыт өте келе әртүрлі генетикалық өзгерістерге ұшырады, нәтижесінде 2019 жылы Ухань қ. ҚХР коронавирустық инфекция адамдар арасында таралды. Әлемде

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2021

коронавирустық инфекцияның кең таралуына байланысты ДДСҰ 2020 жылдың 11 наурызында пандемия жариялады. SARS-CoV-2 коронавирустық инфекциясы әлемнің 200-ге жуық елінде тіркелген және уақыт өте келе әртүрлі генетикалық өзгерістерге ұшырады. Генетикалық өзгерістерге байланысты 2020 жылдың күзінде Ұлыбританияда коронавирустық инфекцияның жаңа штаммы пайда болды. Коронавирустық инфекцияның генетикалық өзгеруін анықтау және оларды ажырату үшін біздің зертханада геномдық толық сиквенс жасау үшін 65 жұп праймер жасалынды. SARS-CoV-2 вирусының маңызды гендерін өңдеуге арналған арнайы праймерлер жасалды. Әзірленген праймерлер SARS-CoV-2 вирустарының гендері туралы толық ақпарат алуға мүмкіндік береді.

Ұсынылған праймерлерді SARS-CoV-2 вирусының толық геномын анықтау және идентификациялау үшін пайдалануға болады

Кілт сөздер: SARS-CoV-2, мутация, ПТР.

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СИНТЕЗ ПРАЙМЕРОВ И НАРАБОТКА ЗНАЧИМЫХ ГЕНОВ В.1.1.7 (АЛЬФА) ВАРИАНТА ВИРУСА SARS-COV-2

Аннотация. Угроза заражения человека коронавирусом стала известна еще в 60-х годах прошлого века. С тех пор зоонозная коронавирусная инфекция распространилась среди людей и в начале 21-го века было две крупные эпидемии. Показатель заражения людей коронавирусной инфекцией быстро распространился и вызвал серьезную обеспокоенность в мире. Коронавирусная инфекция со временем претерпела различные генетические изменения, в результате в 2019 году в городе Ухань, КНР коронавирусная инфекция распространилась среди людей. Из-за широкого распространения коронавирусной инфекции в мире ВОЗ объявила пандемию 11 марта 2020 года. Коронавирусная инфекция SARS-CoV-2 зарегистрирована около в 200 странах мира и со временем претерпела различные генетические изменения. Из-за генетических изменений в Великобритании осенью 2020 года появился новый штамм коронавирусной инфекции. Для определения генетических изменений коронавирусной инфекции и для их различения, в нашей лаборатории было разработано 65 пар праймеров для получения полного геномного сиквенса. Разработаны специальные праймеры для наработки значимых генов вируса SARS-CoV-2. Разработанные праймеры позволяют получить полную информацию о генах вирусов SARS-CoV-2.

Представленные праймеры можно использовать в целях идентификации и наработки полного генома вируса SARS-CoV-2.

Ключевые слова: SARS-CoV-2, мутация, ПЦР.

Introduction. Coronaviruses are a zoonotic and well-studied group of single-stranded RNA viruses of the Coronaviridae family. Coronaviruses (family Coronaviridae) are round enveloped virions with a diameter of 80-220 nm, which contain a single-stranded (+) RNA genome with a size of about 26-32 tn [1].

Since the late 1960s, coronaviruses have been detected in people with mild cold-related symptoms as a causative agent of respiratory diseases [1,2]. Currently, seven different strains

of coronavirus have been described. Of these, 4 coronavirus infections occur in the upper respiratory tract and are characterized by mild symptoms, the remaining 3 coronavirus infections infect the lower respiratory tract and affect a person in severe form, which include: SARS-CoV (*Severe acute respiratory syndrome*), MERS-CoV (*Middle East respiratory syndrome*) and SARS-CoV-2 (*COVID-19*) [3].

In December 2019, a new coronavirus called severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) was discovered in Wuhan, China and caused an epidemic in this area [4]. Due to the spread of coronavirus infection in the world on March 11, 2020, the World Health Organization (WHO) declared COVID-19 a pandemic [5]. As of January 20, a total of 332617707 confirmed cases were registered, resulting in 5551314 deaths. According to WHO data, a total of 1169932 cases, 18338 cases with a fatal outcome have been registered in Kazakhstan to date [6].

All viruses, including SARS-CoV-2 causing COVID-19, undergo genetic changes over time. One of the strains that have undergone genetic changes of COVID-19 is considered to be variant *Alpha B.1.1.7* of the SARS-CoV-2 virus [7]. On May 31, 2021, WHO officially announced the renaming of the British strain to “alpha” [8, 9]. According to WHO, the Alpha coronavirus strain appeared in the UK in September last year. The alpha variant of the coronavirus has an unusually large number of genetic changes, especially in the *Spike* protein that the virus uses when attaching to cells [10]. Three of these mutations have potential biological meanings. These mutations in *Spike* protein include *N501Y*, *P681H* and deletions *H69-V70*, *Y144/145* [11].

Timely accurate diagnosis of infectious diseases occupies an important place in molecular genetic research. In this study, we present a detailed method for developing the complete genome of the Alpha strain of the SARS-CoV-2 coronavirus.

Materials and methods. Clinical samples were obtained from the branch of the Scientific and Practical Center for Sanitary and Epidemiological Expertise and Monitoring of the RSE at the National Center for Public Health of the Ministry of Health of the Republic of Kazakhstan.

Extraction of nucleic acids. Viral RNA was extracted from 140 µl of the sample using *QIAamp Viral mini kits* (Qiagen, Germany) in accordance with the manufacturer's instructions.

Synthesize first – strand cDNA. The reaction was carried out in a *Mastercycler X50s* thermocycler. RT-PCR was performed as follows: 5 µl of extracted RNA was amplified using *SuperScript VILO cDNA Synthesis Kit* and the RT-PCR program was performed under the conditions: 25°C for 10 min; 42°C for 60 min; 85°C for 5 min.

PCR analysis. Amplification was performed in a *Mastercycler X50s* thermocycler using a *Platinum™ SuperFi™ PCR Master Mix (Invitrogen™)*. The master mix of 25 µl, 2X *Platinum™ SuperFi™ PCR Master Mix* – 12,5 µl, 10 µM forward primer – 1,25 µl, 10 µM reverse primer – 1,25 µl, 5X *SuperFi™ GC Enhancer* – 5 µl, *H₂O* – 2 µl, *cDNA* – 3 µl. PCR was performed under the conditions given in Table 1.

Table 1 – Procedure details

Initial denaturation	95°C- 30 сек	1
Denature	95°C- 10 сек	35
Anneal	55°C- 30 сек	
Extend	72°C- 30 сек	
Final extension	72°C- 5 мин	1
	4°C	∞

The following oligonucleotide primers were used for PCR (Table 2):

Table 2 – Developed primers for sequencing the complete genome of the *Alpha* strain of the SARS – CoV-2 coronavirus.

№	Gene	Position	Sequence (5'→3')
1	ORF1a	16-36	Forward primer CTTCCCAGGTAACAAACCAAC
		639-620	Reverse primer TTACGAAGAAGAACCTTGCG
2		111-130	Forward primer TTAGTGCACTCACGCAGTAT
		842-823	Reverse primer GGCCACAGAAGTTGTTATCG
3		767-786	Forward primer GGTGTTACCCGTGAACTCAT
		1467-1448	Reverse primer CCACCCTTACGAAGAATGGT
4		1405-1424	Forward primer TGAGCATAGTCTTGCCGAAT
		2076-2057	Reverse primer CCACCTGTAATGTAGGCCAT
5		1905-1924	Forward primer TTTTCTCCCGCACTCTTGAA
		2643-2624	Reverse primer ATTTGAGCAACATAAGCCC
6		2589-2608	Forward primer AAGCTCCATTGGTTGGTACA
		3262-3243	Reverse primer TGTCTGATTGTCCTCACTGC
7		2872-2891	Forward primer TGAGTTCGCTGTGTTGTGG
		3622-3603	Reverse primer GCCGACAACATGAAGACAGT
8		3429-3450	Forward primer CAGTGGTTGTTAATGCAGCCAA
		4200-4180	Reverse primer ATTTCAGTAGTGCCACCAGCC
9		3724-3744	Forward primer TTTTGGTGCTGACCCATACA
		4401-4382	Reverse primer AGCATTTCTCGAAATTCCA
10		4281-4300	Forward primer TAGAGGAGGCAAAGACAGTG
		4909-4890	Reverse primer ACCATCTAGGTGGAATGTGG
11		4773-4792	Forward primer AAACCATCTCACTTGCTGGT
		5522-5503	Reverse primer TACACACCACGTTCAAGACT
12		5378-5398	Forward primer GGTGAAGCTGCTAACTTTTGT
		6038-6019	Reverse primer AGCTTGCGTTTGATATGGT
13		5811-5832	Forward primer ACGGTGCTTTACTTACAAAGTC
		6558-6539	Reverse primer GCAGCCATTAGATCTGTGTG
14		6372-6391	Forward primer TGGATAATCTTGCTGCGAA
		7119-7100	Reverse primer GAACCAGTACAGTAGGTTGC
15		7005-7024	Forward primer CCGCTGCTTTAGGTGTTTTA
		7691-7672	Reverse primer GTAGTGACAAGTCTCTCGCA
16		7456-7475	Forward primer TGTGCATGTTGTAGACGGTT
		8196-8177	Reverse primer GAATCAACAAACCTTGCCG
17		8013-8032	Forward primer GTGCGGAAGTTGCAGTTAAA
		8715-8696	Reverse primer GTGACACCACCATCAATAGC
18		8534-8554	Forward primer AAGATAGCACTTAAGGGTGGT
		9142-9123	Reverse primer GCCATCCATGAGCACATAAC
19		8808-8827	Forward primer ACAAGCTTGCCCATGATT
		9427-9407	Reverse primer AGCTACAATACCACCAGCTAC
20		9305-9325	Forward primer TTACCAGGAGTTTTCTGTGGT
		10037-10018	Reverse primer TGATAGAGGTTTGTGGTGGT
21		9647-9667	Forward primer TGGATGGTTATGTTACACCT
		10350-10331	Reverse primer GGTGCTTAGGATTGGCTGT
22		10169-10188	Forward primer CCAAGACATGTGATCTGCAC
		10851-10831	Reverse primer GCACACATATCTAAAACGGCA
23		10666-10685	Forward primer TTAGCTTGTTGTACGCTG
		11388-11369	Reverse primer ACTCTCTAGCACCATCATC
24		11226-11245	Forward primer ATATGCCTGCTAGTTGGGTG
		11948-11929	Reverse primer GTAACCTGGACACATTGAGCC
25		11792-11811	Forward primer AAATTGTTGGGTGTTGGTGG
		12438-12419	Reverse primer GGAACACAACCATCTCTTGC
26		12130-12149	Forward primer AGCTTTTGCTACTGCTCAAG
		12799-12780	Reverse primer ACCTCCCTTGTTGTGTTGT
27		12460-12479	Forward primer AACAGCAGCCAACTAATGG
		13181-13162	Reverse primer GACCAGTACCAGTGTGTGTA
28		12891-12910	Forward primer TGGAACCACCTGTAGGTTT
		13542-13523	Reverse primer AGCCCTGTATACGACATCAG

29	ORF1b	13341-13360	Forward primer ACCCTGTGGGTTTTACTT
		14046-14027	Reverse primer AACAAATACCAGCATTTTCGCA
30		13963-13982	Forward primer TACGCCAACTTAGGTGAACG
		14601-14582	Reverse primer TAGATTACCAGAAGCAGCGT
31		14478-14497	Forward primer CCACTTCAGAGAGCTAGGTG
		15190-15171	Reverse primer CTCTAGTGGCGGCTATTGAT
32		14913-14932	Forward primer CCAAGTCATCGTCAACAACC
		15556-15537	Reverse primer CATTAACATTGGCCGTGACA
33		15372-15391	Forward primer GTGTTGTAGCTTGTCACACC
		16030-16011	Reverse primer TAGCTAAAGACACGAACCGT
34		15834-15853	Forward primer ATGTTGGACTGAGACTGACC
		16502-16483	Reverse primer ACTTGTCATTAGCACACAA
35		16374-16393	Forward primer TCCGTATGTTTGCAATGCTC
		17085-17066	Reverse primer TGGTCCCTGGAGGTGAGAAT
36		16743-16762	Forward primer TGGTAAACCTAGACCACCAC
		17492-17473	Reverse primer GGTTCTAGTGTGCCCTTAGT
37		17386-17405	Forward primer TTGAGTGTGTCAATGCCAG
		18043-18024	Reverse primer CAGCTTGAAAGTTGCCACA
38		17832-17851	Forward primer TGTGATTATCACAGGGCT
		18463-18444	Reverse primer GCGGTGGTTTAGCACTAACT
39		18314-18333	Forward primer AGGGGTGTCATGCTACTAGA
		18917-18898	Reverse primer GTCCAGTCAACACGCTTAAC
40		18695-18714	Forward primer CTGCTTCAGACACTTATGCC
		19334-19315	Reverse primer TCAAAAGCTGGTGTGTGGAA
41		19088-19107	Forward primer TCTATGATGCACAGCCTTGT
		19799-19780	Reverse primer TTAGCCCAAAGCTCAAATGC
42		19466-19485	Forward primer GTTGCAATTTAGGTGGTGTCT
		20095-20076	Reverse primer GTTTGGGACCTACAGATGGT
43		19840-19859	Forward primer AATTTGGGTGTGGACATTGC
		20482-20464	Reverse primer ATGAACCTGTTTGCGCATC
44		20077-20096	Forward primer CCATCTGTAGGTCCCAACA
		20676-20657	Reverse primer TTGCCACGCTTGACTAGATT
45		20456-20475	Forward primer TCATAACAGATGCGCAACA
		20166-21147	Reverse primer TTATAGCCACGGAACCTCCA
46		20907-20927	Forward primer TGTTTTAAGACAGTGGTTGCC
		21644-21625	Reverse primer ATGCAGGGGGTAATTGAGTT
47	S	21300-21319	Forward primer ACCACGCGAACAATAGATG
		21921-21901	Reverse primer ACAATAAGTAGGGACTGGGTC
48		21765-21785	Forward primer TACATGTCTCTGGGACCAATG
		22440-22421	Reverse primer AGTGCACAGTCTACAGCATC
49		22345-22364	Forward primer TGCTGCAGCTTATTATGTGG
		23006-22987	Reverse primer CATTACAAGGTGTGCTACCG
50		22849-22868	Forward primer TACAGGCTGCGTTATAGCTT
		23465-23446	Reverse primer CACGCCAAGTAGGAGTAAGT
51		23193-23212	Forward primer ATGGTTTAACAGGCACAGGT
		23914-23894	Reverse primer TTGTGCAAAACTTCTTGGGT
52	S	23498-23517	Forward primer CGTGCAGGCTGTTAATAGG
		24226-24207	Reverse primer AAAGGTCCAACCAGAAGTGA
53		23851-23870	Forward primer AAACCGTGCTTTAACTGGAA
		24571-24552	Reverse primer ACTTTGAAGTCTGCCTGTGA
54		24189-24208	Forward primer CACTGTTAGCGGGTACAATC
		24853-24833	Reverse primer TGAAACAAAGACACCTTCACG
55	ORF3a	24756-24775	Forward primer TGAATTATGTCCCTGCACAA
		25487-25468	Reverse primer GTAGCGCGAACAATACTGA
56		24925-24945	Forward primer TGTGTCTGGTAACTGTGATGT
		25611-25592	Reverse primer GAGTGCTAGTTGCCATCTCT
57		25493-25512	Forward primer CGATACCGATACAAGCCTCA
		26147-26128	Reverse primer GAACCGTCGATTGTGTGAAT
58	ORF3a	25790-25809	Forward primer GCCGTTCCAAAAACCCATTA
		26533-26514	Reverse primer GAACCTGCCATGGCTAAAAT

59	E	26200-26219	Forward primer ACTACTAGCGTGCCTTTGTA
		26922-26903	Reverse primer GAAGCGGTCTGGTCAGAATA
60	M	26795-26814	Forward primer GTGGCTCAGCTACTTCATTG
		27530-27511	Reverse primer AATGGTGAATTGCCCTCGTA
61	ORF6	27426-27445	Forward primer ACTCGCTACTTGAGCTTT
		28096-28077	Reverse primer TTAGAACCAGCCTCATCCAC
62	ORF7a	27874-27893	Forward primer CTTGTCACGCCTAAACGAAC
		28528-28509	Reverse primer GCCAATTTGGTCATCTGGAC
63	ORF8	28288-28307	Forward primer ACCCCAAAATCAGCGAAATG
		29015-28996	Reverse primer TAGTGACAGTTTGGCCTTGT
64	N	28825-28844	Forward primer TCGTTCCTCATCAGTAGTC
		29465-29446	Reverse primer CAGCAGGAAGAAGAGTCACA
65	ORF10	29126-29145	Forward primer AATTTTGGGGACCAGGAAC
		29786-29767	Reverse primer CAGCTCTCCTAGCATTGTT

Horizontal gel electrophoresis was performed in 1.5% agarose solution (*Sigma, USA*), stained with ethidium bromide, in a tris-acetate buffer at a voltage of 100 V/ cm of gel length, for 30 minutes with further detection on a *MiniBIS Pro transilluminator (Israel)*. Visualization and documentation of the results of gel electrophoresis was carried out using the “*GelCapture*” program.

Results and discussion. This study is based on the use of special primers covering the entire genome of the Alpha strain of the *SARS – CoV-2* coronavirus to develop amplified fragments of the nucleotide sequence. To achieve the goal of the work, 65 pairs of specific primers were developed (Table 2) in accordance with the *GenBank* reference sequence: *MN908947.3* [12]. The location of the *SARS – CoV-2* coronavirus genes is shown below (Figure 1).

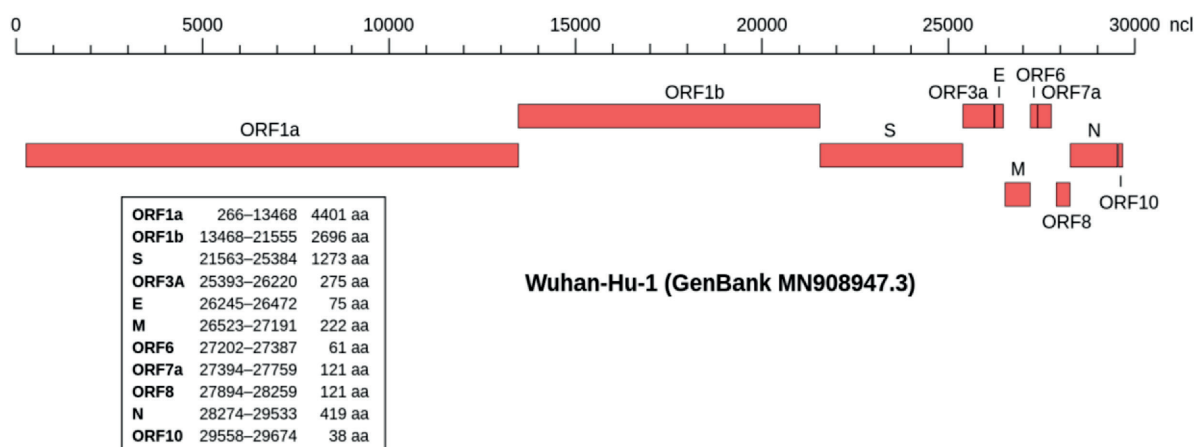


Figure 1 – Organization of the *SARS-CoV-2* coronavirus genome
(<https://www.ncbi.nlm.nih.gov/nuccore/MN908947>)

The *SARS-CoV-2* coronavirus genes were developed using specific primers. The developed *PCR* products are shown in Figures 2 and 3.

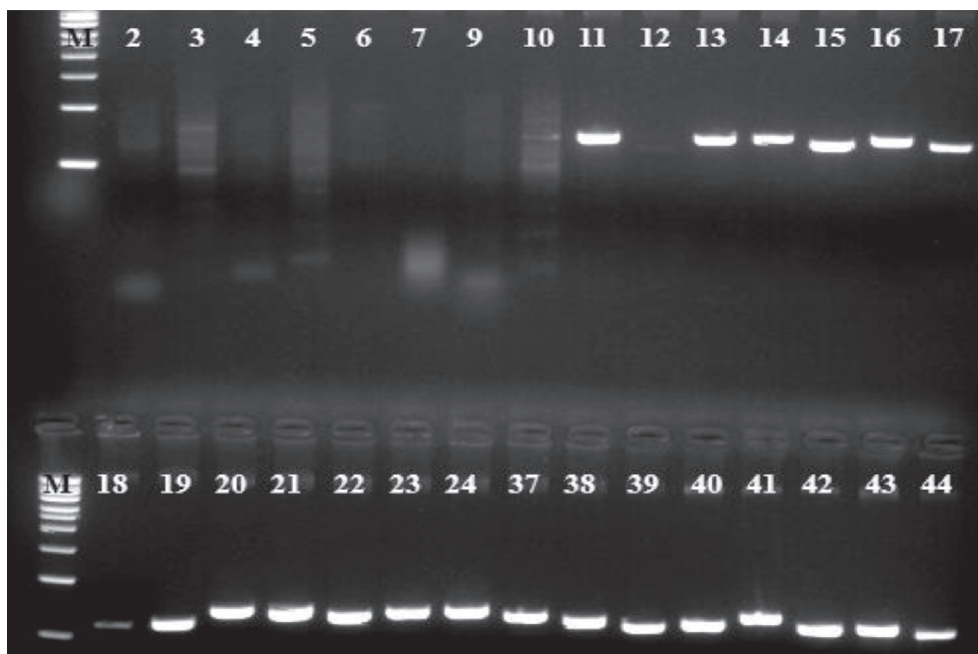


Figure 2 – Electrophoregram of *PCR* products of fragments of the *ORF1a* and *ORF1b* genes of the *Alpha* strain of the *SARS-CoV-2* coronavirus

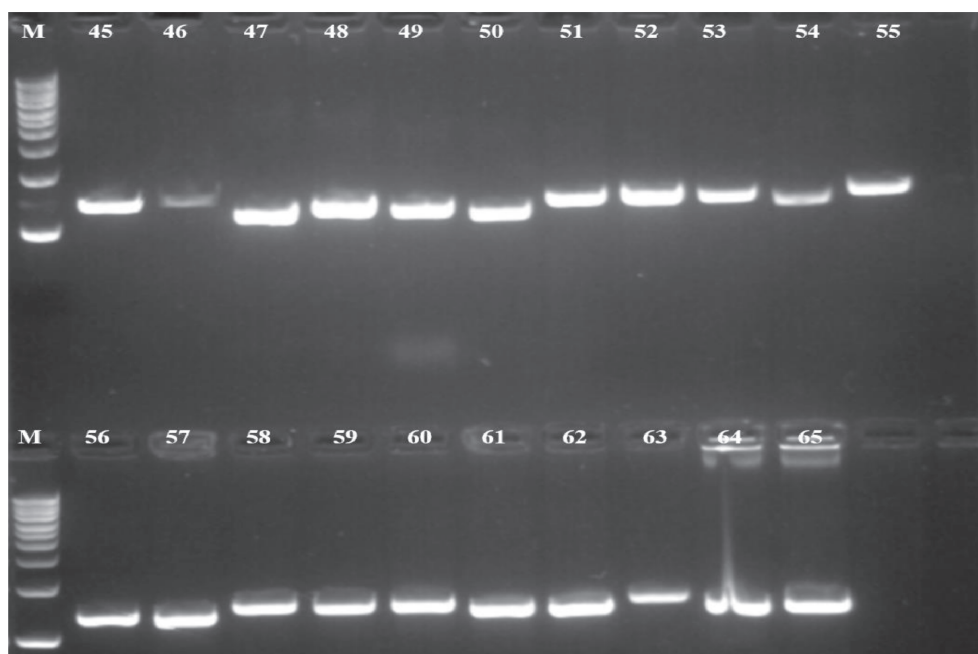


Figure 3 – Electrophoregram of *PCR* products of fragments of genes *S*, *ORF3a*, *E*, *M*, *ORF6*, *ORF7a*, *ORF8*, *N* and *ORF10* of the *Alpha* strain of the *SARS-CoV-2* coronavirus

As can be seen from Figures 2 and 3, *PCR* products of all the genes of the *SARS-CoV-2* virus were developed as *ORF1a*, *ORF1b*, *ORF3a*, *E*, *M*, *ORF6*, *ORF7a*, *ORF8*, *N* and *ORF10*. As a result of the studies in Figure 2, they were amplified with primers (Tab.2) the *ORF1a* and *ORF1b* genes of the *Alpha* strain of the *SARS-CoV-2* coronavirus. Figure 3 shows the accumulated *PCR* products of the genes *S*, *ORF3a*, *E*, *M*, *ORF6*, *ORF7a*, *ORF8*, *N* and *ORF10* of the *Alpha* strain

of the SARS-CoV-2 coronavirus. As a result of the work carried out, PCR products of the above-mentioned genes of the *Alpha* variant of the SARS-CoV-2 coronavirus have been developed, which are prepared for full-genome sequencing.

Conclusion. Coronaviruses are a well-studied group of single-stranded (+) RNA viruses of the Coronaviridae family. The facts of human infection with coronavirus have become known since the 60s of the last century. Before the global SARS-CoV-2 pandemic, there was an epidemic of 2 coronavirus cases worldwide: SARS-CoV (severe acute respiratory syndrome) and MERS-CoV (*Middle East respiratory syndrome*). According to WHO, a total of 332617707 cases, 5551314 cases of death have been registered in the world today.

The molecular genetic study of the MERS-CoV-2 coronavirus is an important measure for assessing the prevalence of the disease and analyzing genomic mutations. As a result of such studies, a new strain of coronavirus was identified in the UK in the autumn of 2020. Depending on the place of origin, this strain has been called the British strain. On May 31, 2021, WHO assigned the name “alpha” to this strain. Using PCR, the genes of the Alpha strain of the MERS-CoV-2 coronavirus were developed using specially developed primers. In the future, as a result of these works, full-genome sequencing of the Alpha strain of the SARS-CoV-2 coronavirus is expected.

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