






SEQUENCING AND PHYLOGENETIC ANALYSIS OF THE ORF3A GENE VARIANT B.1.1.7 SARS-COV-2 VIRUS

B.S. Usserbayev *, N.S. Kozhabergenov , A.M. Melisbek ,
 M.Zh. Shirinbekov , A.I. Tagayev , Ye.D. Burashev 

«Research Institute for Biological Safety Problems», MH RK, Gvardeyskiy uts
 ussербayev.bekbolat@mail.ru

Abstract. The problem of «new» infectious diseases has now acquired particular relevance for national and international health systems. One of these infections is the SARS-CoV-2 coronavirus. Due to the rapid spread of a new type of coronavirus in the world, molecular genetic studies of this virus are of great importance. This will help to understand the nature of the virus and develop antiviral drugs to prevent the disease. The aim is to identify a missense mutation in the ORF3 gene of the studied coronavirus strain *SARS-CoV-2/human/KAZ/Britain*. This paper presents the results of the development and sequencing of the ORF3 gene of the coronavirus strain *SARS-Cov-2/human/KAZ/Britain*. For the development and sequencing by the Sanger method of the ORF3 gene, 4 pairs of primers were developed, with an overlap of 100-150 bp. The changes in the ORF3 gene compared to other strains whose data were obtained from the GISAID database were analyzed. As a result, the obtained amino acid sequences of the ORF3a gene of variant B.1.1.7 were compared with the reference strain HCoV-19/Wuhan/Hu-1/2019, where one mutation was determined at position 149: W (tryptophan) → L (leucine). The phylogenetic affiliation of the studied virus strain was also determined, where the genetic distance between the studied strains of the SARS-CoV-2 virus was determined. Thus, the greatest similarity with the studied strain were:

HCoV-19/Wuhan/Hu-1/2019,
HCoV-19/England/MILK-9E05B3/2020,
HCoV-19/Japan/IC-0446/2020,
HCoV-19/Germany/NI-IOV-MHH15/2020,
 and the strain *HCoV-19/Argentina/PAIS-F0418/*
2021 significantly distanced itself from the *SARS-CoV-2/human/KAZ/Britain* strain.

Keywords: SARS-CoV-2; mutation; PCR; primer; sequencing

Introduction

At the end of 2019, the republican office of the World Health Organization (WHO) in the People's Republic of China (PRC) was informed by the health authorities of the city of Wuhan about the registration of cases of pneumonia of unknown etiology [1, 2]. At the beginning of January 2020, 44 patients with pneumonia of unknown etiology were registered in China. The causative agent of the infection was not detected, and therefore WHO requested additional information from the national authorities of China to assess the risk [3]. Epidemiological studies conducted by the Chinese Center for Disease Control and Prevention (CDC China) and the Wuhan City Health Authority showed that the outbreak was associated with the seafood market, and with the help of genome-wide sequencing, a new (2019-nCoV) type of

coronavirus was identified, whose genome was about 29.9 bp. (GenBank NC_045512.2) [4, 5]. At the 5' end of the genome, 2019-nCoV encodes a replicase gene containing two open reading frames (ORF) ORF1 and ORF1b. The ORF gene contains 15 nonstructural proteins (NSP), including from NSP 1 to NCP10 and from NSP 12 to NSP 16 [5, 6]. At the 3' end, the genome contains the genes S, ORF3a, E, membrane protein (M), ORF6, ORF7a, ORF7b, ORF8, N and protein ORF10 [7, 8]. Among the proteins listed above, ORF3a, ORF8 and ORF10 are unique for a new coronavirus infection [9].

In this work, attention is focused on the ORF3a protein, since it participates in important aspects of the pathogenicity of the virus [10] and includes various functional domains depending on virulence, infectivity [11]. The ORF3a protein is located between the loci of the S and E genes, which forms the N-terminal transmembrane and C-terminal domains. The ORF3a - protein is viroporin, a viral transmembrane protein that exhibits the properties of ion channels in cell membranes [12, 13]. The ORF3a protein interacts with caveolin, which regulates various phases of the viral cycle [14]. It should be noted that a significant reaction of CD4+ and CD8+T cells to SARS-CoV-2 in infected people was directed against ORF3a [15].

The ORF3a protein was initially found in the coronavirus family, and then described under various names, such as protein X1 [16], protein 3a [17], U274 [18] and ORF3a [14]. At the beginning of the COVID-19 pandemic in 2019, we learned about the pathogenic property of the ORF3a gene of the SARS-CoV-2 virus [19]. Some studies have shown that many recovering patients with atypical pneumonia produce antibodies to ORF3a and the serum of patients with COVID-19 also demonstrates a high level of IgG and IgA reactivity to ORF3a, in addition to structural proteins [20]. The analysis of nucleotide and protein sequences of infection of a new variant of the coronavirus SARS-CoV-2 provides important data on the biology of the virus. Therefore, molecular genetic studies are important for monitoring the evolution of the virus.

The aim of this work is to identify changes in the ORF3a gene of the studied coronavirus strain SARS-CoV-2/human/KAZ/Britain.

Materials and methods

Research material. The coronavirus strain SARS-CoV-2/human/KAZ/Britain from the laboratory of the collection of microorganisms of the Research Institute of Biological Safety Problems was used as the object of the study.

Isolation of RNA. Virus RNA was extracted from a virus-containing liquid with the QIAamp Viral RNA Mini kit (Qiagen, Germany) in accordance with the manufacturer's instructions in the BSL-3 laboratory.

cDNA synthesis. Reverse transcription (OT) was performed using the SuperScript VILO cDNA Synthesis Kit (Invitrogen, Thermo Fisher Scientific, USA) in a Mastercycler X50s thermal cycler at 25°C for 10 min; 42°C for 60 min; 85°C for 5 min.

Selection and synthesis of primers. The search and selection of specific primers for the development and sequencing of the ORF3a gene of the SARS-CoV-2 virus was carried out using the online program Primer-BLAST (<http://www.ncbi.nlm.nih.gov/tools/primer-blast>). The synthesis of oligonucleotides was carried out on an automatic DNA/RNA Synthesizer H16 oligonucleotide synthesizer (K&A Laborgeraete, Germany) by the amidophosphite method.

Setting up PCR. Amplification was performed on a Mastercycler X50s thermal cycler using a Platinum Super Fi PCR Master Mix kit (Invitrogen, Thermo Fisher Scientific, USA) in accordance with the manufacturer's instructions. The operating time of PCR products was carried out according to the following temperature regime: initial denaturation 95°C-0.5 min; followed by 35 cycles of amplification at denaturation 95°C-0.1 min, annealing 57°C-0.5 min, elongation 72°C-0.5 min; final elongation 72°C-5 min.

Gel electrophoresis. Horizontal gel electrophoresis was performed in 1.5% agarose gel (Sigma, USA), stained with ethidium bromide, in a tris-acetate buffer at a voltage of 100 volts / cm of gel length, for 30 minutes with further detection on the MiniBIS Pro transilluminator (DNR Bio Imaging Systems, Israel). Visualization and documentation of the results of gel electrophoresis was carried out using the «GelCapture» program 1 kb DNA Ladder ready-to-use (Bioron, Germany) was used as a marker of molecular weights.

Purification of the PCR product. The PCR purification of the product was carried out with the GeneJET PCR Purification Kit (Thermo Fisher Scientific, USA) according to the manufacturer's instructions.

Sequencing by the Sanger method. Sequencing of the ORF3a gene of the SARS-CoV-2 virus after purification of the PCR product was carried out using cyclic sequencing by the Sanger method with a set of AB BigDye Terminator v3.1 Cycle Sequencing Kit (Thermo Fisher Scientific, USA) and sequencing primers (Table 1), which were used at the production stage. The products were purified with a set of BigDye Xterminator (Thermo Fisher Scientific, USA) and sequenced using a 3130 XL genetic analyzer (Applied Biosystems).

Bioinformatic analysis and algorithm. After sequencing, the obtained nucleotide sequence data were processed using the Sequencher v.5.4 program (Gene Codes Corporation, USA). Comparison of the amino acid sequence of the ORF3 gene with the reference strain was carried out using the Protein BLAST program (<https://blast.ncbi.nlm.nih.gov>) based on NCBI. Multiple alignment, phylogenetic analysis and calculation of genetic distances of the gene were carried out using the MEGA 11 program [21, 22].

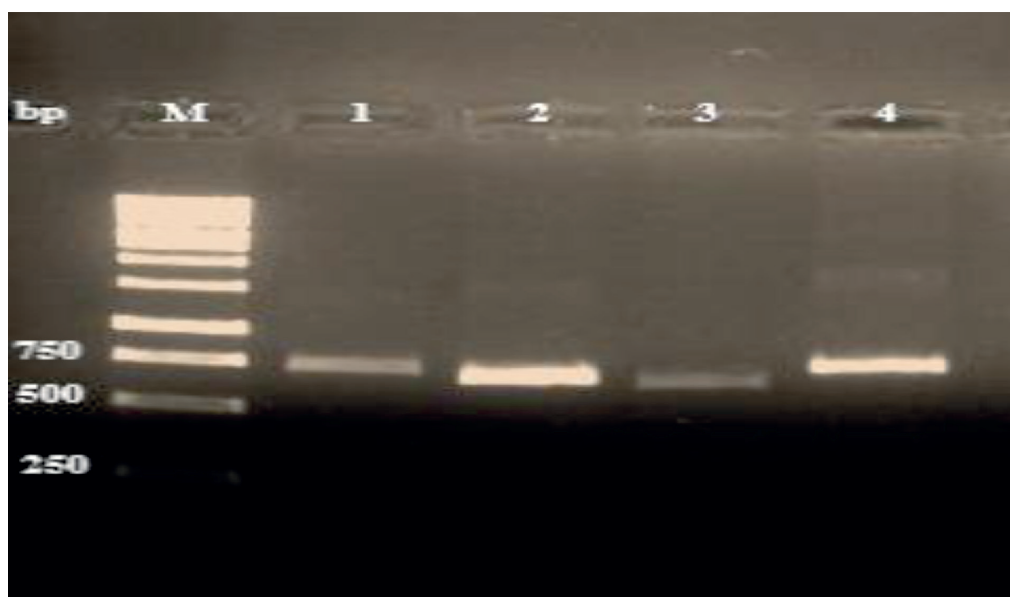
The main results of the research. The sequence of nucleotides of specific primers was selected based on the ORF3 gene of the reference strain *Wuhan-Hu-1*. Primers were selected in such a way that each pair of primers overlapped each other. As a result, 4 pairs of sequencing primers were selected (Table.1) to develop the complete ORF3a gene of the SARS-CoV-2 virus with an overlap of about 100 bp.

Table 1 – Parameters of sequencing primers

| Nº | Sequence 5' – 3' | Length | Tm | GC% | Position | Product size, bp |
|----|---------------------------------------|--------|-------|-------|-------------|------------------|
| f | Forward primer TGA CTTATG TCCCTGCACAA | 20 | 56.76 | 45.00 | 24756-24775 | 731 |
| r | Reverse primer GTAGCGCGAACAAAATCTGA | 20 | 56.85 | 45.00 | 25487-25468 | |
| f | Forward primer TGTGTCTGGTAACTGTGATGT | 21 | 56.88 | 42.86 | 24925-24945 | 686 |
| r | Reverse primer GAGTGCTAGTTGCCATCTCT | 20 | 57.02 | 50.00 | 25611-25592 | |
| f | Forward primer CGATACCGATACAAGCCTCA | 20 | 56.92 | 50.00 | 25493-25512 | 654 |
| r | Reverse primer GAACCGTCGATTGTGTGAAT | 20 | 56.47 | 45.00 | 26147-26128 | |
| f | Forward primer GCCGTTCCAAAACCCATTA | 20 | 56.89 | 45.00 | 25790-25809 | 743 |
| r | Reverse primer GAACCTGCCATGGCTAAAAT | 20 | 56.35 | 45.00 | 26533-26514 | |

Selected primers (Table.1) allow to fully develop the ORF3a gene of the SARS-CoV-2 virus, the length of which is 825 bp [19].

Thus, with the selected primers (Table 1), 4 fragments of the ORF3a gene of the SARS-CoV-2 virus were developed (Fig.1).



M – marker; 1–4 test samples

Figure 1 – Electrophoregram of PCR products of the *ORF3a* gene strain *SARS-CoV-2/human/KAZ/Britain*.

As can be seen from Figure 1, after carrying out standard PCR with a set of specific primers (Table 1), fragments of PCR products were developed in appropriate sizes.

As a result, the amino acid sequences of the obtained *ORF3a* gene were compared with the reference strain, where one mutation was detected at position 149 (Fig.2).

[Download](#) [Graphics](#)

unnamed protein product

Sequence ID: **Query_16105** Length: **275** Number of Matches: **1**

Range 1: 1 to 275 [Graphics](#)

[Next Match](#) [Previous Match](#)

| Score | Expect | Method | Identities | Positives | Gaps |
|----------------|---|------------------------------|--------------|--------------|-----------|
| 565 bits(1456) | 0.0 | Compositional matrix adjust. | 274/275(99%) | 274/275(99%) | 0/275(0%) |
| Query 1 | MDLFMRIFTIGTVTLKQGEIKDATPSDFVRATATIPQASLPFGWLIVGVALLAVFQSAS | 60 | | | |
| Sbjct 1 | | 60 | | | |
| Query 61 | KIITLKKRWQLALSKGVHFCNLLLLFVTVYSHLLLVAAGLEAPFLYLYALVYFLQSINF | 120 | | | |
| Sbjct 61 | | 120 | | | |
| Query 121 | VRIIMRLWLCWKCRSKNPLLYDANYFLCLHTNCYDYCIPYNSVTSSIVITSGDGTTSPIIS | 180 | | | |
| Sbjct 121 |W | 180 | | | |
| Query 181 | EHDYQIGGYTEKWESGVKDCVVLHSYFTSDYYQLYSTQLSTDTGVEHVTFIYNKIVDEP | 240 | | | |
| Sbjct 181 | | 240 | | | |
| Query 241 | EEHVQIHTIDGSSGVVNPVMEPIYDEPTTTTSVPL | 275 | | | |
| Sbjct 241 | | 275 | | | |

Figure 2 – Comparison of the amino acid sequence of the analyzed *ORF3a* gene with the reference strain Wuhan-Hu-1.

As a result of the data obtained, a variation was determined that led to a corresponding change in the amino acid from W (Tryptophan) to L (Leucine) at the amino acid position 149.

Next, a phylogenetic analysis of the studied viral strain by the ORF3a gene was performed in order to determine the most phylogenetically close to it viral strains of SARS-CoV-2 (Fig.3). The analysis used 12 other global strains obtained from the database of the Global Initiative for the Exchange of All Influenza Data (GISAID) (<https://www.gisaid.org>).

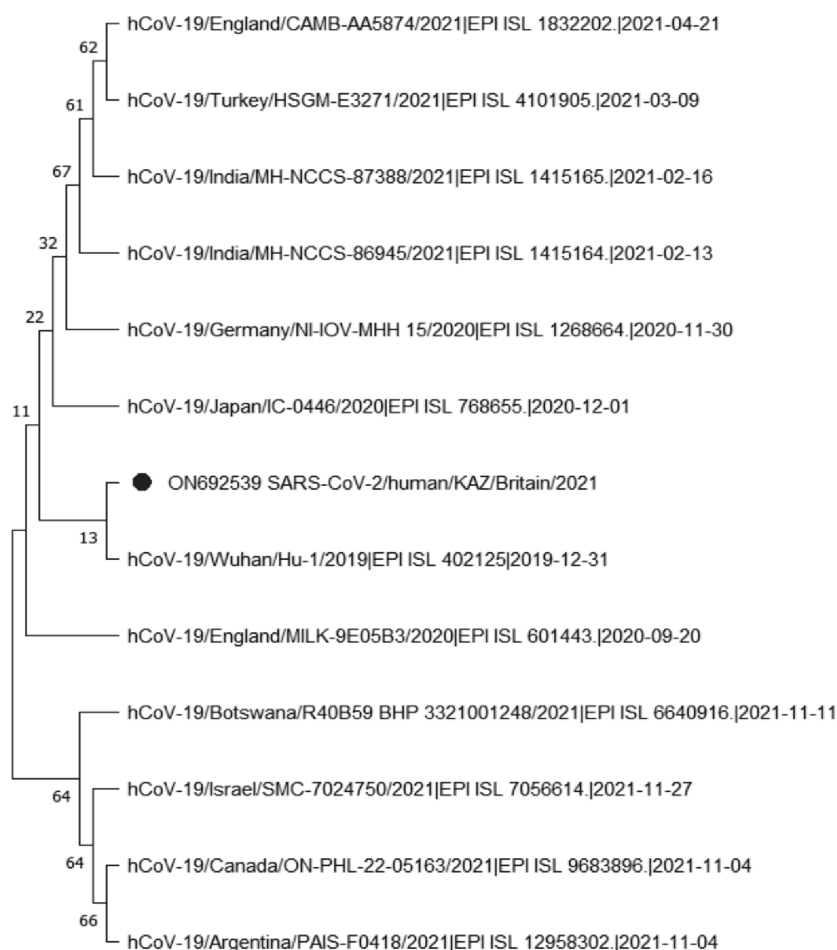


Figure 3 – Phylogenetic tree of the *SARS-CoV-2/human/KAZ/Britain* strain and other *SARS-CoV-2* strains by the ORF3a gene.

Phylogenetic analysis showed that the viral strains *SARS-CoV-2/human/KAZ/Britain* and *HCoV-19/Wuhan/Hu-1/2019* are identical to each other in this part of the genome.

When analyzing the assessment of the evolutionary divergence between the sequences of the ORF3 gene of the strain under study and the data (Fig.2) of GISAID, the number of basic differences between the nucleotide sequences of the gene was determined (Table 2).

Table 2 – Genetic differentiation of the ORF3a gene of the *SARS-CoV2/human/KAZ/Britain* strain and other strains obtained from the GISAID database

| № | Name of strains | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 |
|----|---|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|
| 1 | SARS-CoV-2/human/KAZ/ Britain | | 0,00120 | 0,00120 | 0,00120 | 0,00120 | 0,00124 | 0,00124 | 0,00162 | 0,00216 | 0,00178 | 0,00178 | 0,00178 | 0,00255 |
| 2 | hCoV-19/Wuhan/Hu-1/2019 | 0,00121 | | 0,00000 | 0,00000 | 0,00000 | 0,00000 | 0,00000 | 0,00115 | 0,00179 | 0,00130 | 0,00130 | 0,00130 | 0,00215 |
| 3 | hCoV-19/England/ MILK-9E05B3/2020 | 0,00121 | 0,00000 | | 0,00000 | 0,00000 | 0,00000 | 0,00000 | 0,00115 | 0,00179 | 0,00130 | 0,00130 | 0,00130 | 0,00215 |
| 4 | hCoV-19/Japan/IC-0446/2020 | 0,00121 | 0,00000 | 0,00000 | | 0,00000 | 0,00000 | 0,00000 | 0,00115 | 0,00179 | 0,00130 | 0,00130 | 0,00130 | 0,00215 |
| 5 | hCoV-19/Germany/ NI-IOV-MHH15/2020 | 0,00121 | 0,00000 | 0,00000 | 0,00000 | | 0,00000 | 0,00000 | 0,00115 | 0,00179 | 0,00130 | 0,00130 | 0,00130 | 0,00215 |
| 6 | hCoV-19/India/ MH-NCCS86945/2021 | 0,00126 | 0,00000 | 0,00000 | 0,00000 | 0,00000 | | 0,00000 | 0,00000 | 0,00132 | 0,00134 | 0,00134 | 0,00134 | 0,00223 |
| 7 | hCoV-19/India/ MH-NCCS87388/2021 | 0,00126 | 0,00000 | 0,00000 | 0,00000 | 0,00000 | 0,00000 | | 0,00000 | 0,00132 | 0,00134 | 0,00134 | 0,00134 | 0,00223 |
| 8 | hCoV-19/England/ CAMB-AA5874/2021 | 0,00243 | 0,00121 | 0,00121 | 0,00121 | 0,00121 | 0,00000 | 0,00000 | | 0,00127 | 0,00178 | 0,00178 | 0,00178 | 0,00244 |
| 9 | hCoV-19/Turkey/ HSGM-E3271/2021 | 0,00365 | 0,00243 | 0,00243 | 0,00243 | 0,00243 | 0,00126 | 0,00126 | 0,00121 | | 0,00221 | 0,00221 | 0,00221 | 0,00275 |
| 10 | hCoV-19/ Botswana /R40659 BHP 3321001248 /2021 | 0,00243 | 0,00121 | 0,00121 | 0,00121 | 0,00121 | 0,00125 | 0,00125 | 0,00242 | 0,00364 | | 0,00000 | 0,00000 | 0,00175 |
| 11 | hCoV-19/Israel/ SMC-7024750/2021 | 0,00243 | 0,00121 | 0,00121 | 0,00121 | 0,00121 | 0,00125 | 0,00125 | 0,00242 | 0,00364 | 0,00000 | | 0,00000 | 0,00175 |
| 12 | hCoV-19/Canada/ ON-PHL-22-05163/2021 | 0,00243 | 0,00121 | 0,00121 | 0,00121 | 0,00121 | 0,00125 | 0,00125 | 0,00242 | 0,00364 | 0,00000 | 0,00000 | | 0,00175 |
| 13 | hCoV-19/Argentina/ PAIS-F0418/2021 | 0,00486 | 0,00364 | 0,00364 | 0,00364 | 0,00364 | 0,00377 | 0,00377 | 0,00485 | 0,00609 | 0,00242 | 0,00242 | 0,00242 | |

From the data obtained, it can be seen that the nucleotide sequence of the ORF3a gene showed the greatest similarity between the strains *SARS-CoV-2/human/KAZ/Britain* and *HCoV-19/Wuhan/Hu-1/2019*, *HCoV-19/England/MILK-9E05B3/2020*, *HCoV-19/Japan/IC-0446/2020*, *HCoV-19/Germany/NI-IOV-MHH15/2020*, the divergence coefficient of which is $D \geq 0.00121$. Pairwise values of genetic differentiation (Table 2) based on the ORF3a gene showed that the strain *SARS-CoV-2/human/KAZ/Britain* significantly distances itself from the strain *HCoV-19/Argentina/PAIS-F0418/2021* ($D \geq 0.00486$).

Discussion of the results obtained. According to data from *GISAID*, there are currently 12 different clades of the *SARS-CoV-2* virus in the world (www.gisaid.org). A significant place for the evolutionary control and differentiation of mutational variability of microorganisms is occupied by molecular genetic studies. Sanger sequencing is a reference sequencing method in which the probability of incorrect reading is very low compared to other next-generation sequencing methods.

In this research paper, we presented data on the sequencing of the ORF3a gene of the *SARS-CoV-2* virus. For these purposes, we have developed 4 pairs of sequencing primers, which together develop the whole ORF3a gene of the *SARS-CoV-2* coronavirus (Fig.1). During the research, the optimal parameters of the PCR formulation were determined, which corresponds to the following values: the annealing temperature is 57°C, and the concentration of each primer is 400 mM in the reaction mixture. According to the WHO classification, mutations in the ORF3a gene were detected in the groups of virus of interest (VOI) and virus of concern (VOC) (Table 3) [19].

Table 3 – Brief description of new mutant variants for the ORF3a gene of the *SARS-CoV-2* virus [19]

| Вирусные варианты | Pango lineage | First detection and location detection | ORF3a mutations |
|-------------------------|------------------|--|------------------|
| Virus of Interest (VOI) | | | |
| <i>Lambda</i> | C.37 | Peru, December - 2020 | No |
| <i>Mu</i> | B.1.621 | Colombia, January - 2021 | Q57H, del256/257 |
| <i>Epsilon</i> | B.1.427 | California, USA, June - 2020 | Q57H |
| Virus of Concern (VOC) | | | |
| <i>Alpha</i> | B.1.1.7 | United Kingdom, September - 2020 | No |
| <i>Beta</i> | B.1.351 | South Africa, May - 2020 | Q57H, S171L |
| <i>Gamma</i> | P.1 | Brazil, November - 2020 | S253P, D155Y |
| <i>Delta</i> | B.1.617.2 | India, October - 2020 | S26L |
| <i>Omicron</i> | BA.1/B.1.1.529 | South Africa November - 2021 | No |
| | BA.2/B.1.1.529.2 | South Africa November - 2021 | T223I |
| | BA.3/B.1.1.529.3 | South Africa November - 2021 | T223I |

As can be seen from Table 3, the Q57H mutation is present in the «Epsilon» and Q57H/S171L «Beta» variants [23]. The appearance of the Q57H mutation is associated with the 4th wave of the revival of *SARS-CoV-2* in Hong Kong [24]. In the «Gamma» variant, only a single S253P change or combined S253P/D155Y mutations were detected. Among the sarbecovirus group, D155 and S253 residues are considered to be the most conservative regions, and these mutations cause a change in the activity of the ORF3a protein [19]. The S26L mutation found in the «Delta» variant does not affect the nature of the virus, since this region is very diverse among the sarbecovirus family [25]. Currently, there is no evidence that the T223I mutation found in the «Omicron» variant affects the virulence of the virus [26, 27].

In this work, we identified a change in one amino acid in the ORF3a gene of the *SARS-CoV-2/human/KAZ/Britain* strain. Davarynejad et.al [28] showed that replacing just one

amino acid in one protein is crucial for cell division. This requires regular genomic and epidemiological studies to identify new variants of viral pathogens. Since the amino acid sequence was detected, it was found only in the strain under study compared to other strains of *SARS-CoV-2* variants (https://cov-lineages.org/global_report_B.1.1.7.html). This identified amino acid sequence requires effective research in the future, since it is unknown whether this affects the virulence and functional nature of the virus.

Phylogenetic analysis showed that the studied strain is close to the reference strain compared to other variants of *SARS-CoV-2* strains.

However, social distancing of the population, non-compliance with personal protective measures and low vaccination rates also play a significant role in the spread of coronavirus infection. Thus, molecular genetic analysis further activates the need for genomic studies to identify new changes in the genes of *SARS-CoV-2* variants.

Conclusion. As a result of research work, four pairs of primers were selected for the development and sequencing by the Sanger method of the ORF3a gene of variant B.1.1.7, with a 100-150 bp overlap. The results of the study showed that PCR products fully correspond to the length of the developed primers.

The obtained amino acid sequences of the ORF3a gene were compared with a reference strain (GenBank inventory number NC_045512.2) where one mutation was determined at position 149: W→L. Comparative genetic analysis showed that the nucleotide sequences of the ORF3a gene of the coronavirus strain *SARS-CoV-2/human/KAZ/Britain* are highly identical to the ORF3a gene of the *HCoV-19/Wuhan/Hu-1/2019* strain.

The analysis of the nucleotide sequence of the ORF3a gene of the *SARS-CoV-2/human/KAZ/Britain* strain showed the greatest similarity with *HCoV-19/Wuhan/Hu-1/2019*, *HCoV-19/England/MILK-9E05B3/2020*, *HCoV-19/Japan/IC-0446/2020*, *HCoV-19/Germany/NI-IOV-MHH15/2020*. The genetic distances revealed by the nucleotide sequences of the ORF3a gene are significantly distanced from *HCoV-19/Argentina/PAIS-F0418/2021*.

The work was carried out within the framework of the grant funding project AP09058338 «Study of antiviral activity of drugs in relation to the SARS-COV-2 virus in vitro and molecular epidemiological analysis of circulating COVID-19 strains».

The authors of this article declare that there is no conflict of interest.

References

- 1 Lu H., Stratton C.W., Tang Y.W. Outbreak of pneumonia of unknown etiology in Wuhan, China: The mystery and the miracle. // J Med Virol. – 2020. 92(4). - P.401-402. doi:10.1002/jmv.25678
- 2 Lam S., Lombardi A., Ouanounou A. COVID-19: A review of the proposed pharmacological treatments. // Eur J Pharmacol. – 2020. - 886 (5) 173451. doi:10.1016/j.ejphar.2020.173451
- 3 COVID-19 – China. Pneumonia of unknown cause – China (who.int)
- 4 Zhu N., Zhang D., Wang W., Li X., Yang B., Song J., Zhao X., Huang B., Shi W., Lu R., Niu P., Zhan F., Ma X., Wang D., Xu W., Wu G., Gao G.F., Tan W; China Novel Coronavirus Investigating and Research Team. A Novel Coronavirus from Patients with Pneumonia in China, 2019. // N Engl J Med. – 2020. 382(8). – P. 727-733. doi:10.1056/NEJMoa2001017
- 5 Wu F., Zhao S., Yu B., Chen Y.M., Wang W., Song Z.G., Hu Y., Tao Z.W., Tian J.H., Pei Y.Y., Yuan M.L., Zhang Y.L., Dai F.H., Liu Y., Wang Q.M., Zheng J.J., Xu L., Holmes E.C., Zhang Y.Z. A new coronavirus associated with human respiratory disease in China. // Nature. – 2020. 579 (7798). – P. 265-269. doi:10.1038/s41586-020-2202-3
- 6 Emam M., Oweda M., Antunes A., El-Hadidi M. Positive selection as a key player for SARS-CoV-2 pathogenicity: Insights into ORF1ab, S and E genes. // Virus Res. – 2021. 302:198472. doi:10.1016/j.virusres.2021.198472
- 7 SeyedAlinaghi S., Mirzapour P., Dadras O. et al. Characterization of SARS-CoV-2 different variants and related morbidity and mortality: a systematic review. // Eur J Med Res. - 2021. 26(1). 51. doi:10.1186/s40001-021-00524-8

- 8 Wu A., Peng Y., Huang B., Ding X., Wang X., Niu P., Meng J., Zhu Z., Zhang Z., Wang J., Sheng J., Quan L., Xia Z., Tan W., Cheng G., Jiang T. Genome Composition and Divergence of the Novel Coronavirus (2019-nCoV) Originating in China. // *Cell Host Microbe*. – 2020. 27(3). – P.325-328. doi:10.1016/j.chom.2020.02.001
- 9 Gordon, D. E., Jang, G. M., Bouhaddou, M., Xu, J., Obernier, K., White, K. M., et al. A SARS-CoV-2 protein interaction map reveals targets for drug repurposing. // *Nature*. – 2020. 583. – P. 459–468. doi:10.1038/s41586-020-2286-9
- 10 Ren, Y., Shu, T., Wu, D. et al. The ORF3a protein of SARS-CoV-2 induces apoptosis in cells. // *Cell Mol Immunol*. – 2020. 17. – P. 881–883. doi:10.1038/s41423-020-0485-9
- 11 Issa E, Merhi G, Panossian B, Salloum T, Tokajian S. SARS-CoV-2 and ORF3a: nonsynonymous mutations, functional domains, and viral pathogenesis.” // *mSystems*. – 2020. 5(3). doi:10.1128/mSystems.00266-20
- 12 Scott C., Griffin S. Viroporins: structure, function and potential as antiviral targets. // *J Gen Virol*. – 2015 96(8). – P. 2000-2027. <https://doi.org/10.1099/vir.0.000201>
- 13 Bianchi M., Borsetti A., Ciccozzi M., Pascarella S. SARS-CoV-2 ORF3a: Mutability and function. // *Int J Biol Macromol*. – 2021. 170. – P. 820-826. doi: 10.1016/j.ijbiomac.2020.12.142
- 14 Padhan K., Tanwar C., Hussain A., Hui P.Y., Lee M.Y., Cheung C.Y., Peiris J.S.M., Jameel S. Severe acute respiratory syndrome coronavirus Orf3a protein interacts with caveolin. // *J Gen Virol*. – 2007. 88(11). – P. 3067-3077. <https://doi.org/10.1099/vir.0.82856-0>
- 15 Grifoni A., Weiskopf D., Ramirez S.I., Mateus J., Dan J.M., Moderbacher C.R., Rawlings S.A., Sutherland A., Premkumar L., Jadi R.S., Marrama D., de Silva A.M., Frazier A., Carlin A.F., Greenbaum J.A., Peters B., Krammer F., Smith D.M., Crotty S., Sette A. Targets of T Cell Responses to SARS-CoV-2 Coronavirus in Humans with COVID-19 Disease and Unexposed Individuals. // *Cell*. – 2020. 181(7). – P.1489-1501. doi:10.1016/j.cell.2020.05.015
- 16 Rota P.A., Oberste M.S., Monroe S.S., Nix W.A., Campagnoli R., Icenogle J.P., Peñaranda S., Bankamp B., Maher K., Chen M.H., Tong S., Tamin A., Lowe L., Frace M., DeRisi J.L., Chen Q., Wang D., Erdman D.D., Peret T.C., Burns C., Ksiazek T.G., Rollin P.E., Sanchez A., Liffick S., Holloway B., Limor J., McCaustland K., Olsen-Rasmussen M., Fouchier R., Günther S., Osterhaus A.D., Drosten C., Pallansch M.A., Anderson L.J., Bellini W.J. Characterization of a novel coronavirus associated with severe acute respiratory syndrome. // *Science*. – 2003. 300(5624). –P.1394-9. doi:10.1126/science.1085952
- 17 Marra M.A., Jones S.J., Astell C.R., Holt R.A., Brooks-Wilson A., Butterfield Y.S., Khattra J., Asano J.K., Barber S.A., Chan S.Y., Cloutier A., Coughlin S.M., Freeman D., Girn N., Griffith O.L., Leach S.R., Mayo M., McDonald H., Montgomery S.B., Pandoh P.K., Petrescu A.S., Robertson A.G., Schein J.E., Siddiqui A., Smailus D.E., Stott J.M., Yang G.S., Plummer F., Andonov A., Artsob H., Bastien N., Bernard K., Booth T.F., Bowness D., Czub M., Drebot M., Fernando L., Flick R., Garbutt M., Gray M., Grolla A., Jones S., Feldmann H., Meyers A., Kabani A., Li Y., Normand S., Stroher U., Tipples G.A., Tyler S., Vogrig R., Ward D., Watson B., Brunham R.C., Krajden M., Petric M., Skowronski D.M., Upton C., Roper R.L.. The Genome sequence of the SARS-associated coronavirus. // *Science*. – 2003. 300(5624). –P.1399-404. doi:10.1126/science.1085953
- 18 Tan Y.J., Teng E., Shen S., Tan T.H., Goh P.Y., Fielding B.C., Ooi E.E., Tan H.C., Lim S.G., Hong W. A novel severe acute respiratory syndrome coronavirus protein, U274, is transported to the cell surface and undergoes endocytosis. // *J Virol*. – 2004. 78(13). – P.6723-34. doi:10.1128/JVI.78.13.6723-6734.2004
- 19 Zhang J., Ejikemeuwa A., Gerzanich V., Nasr M., Tang Q., Simard J.M., Zhao R.Y. Understanding the Role of SARS-CoV-2 ORF3a in Viral Pathogenesis and COVID-19. // *Front Microbiol*. – 2022. 13:854567. doi:10.3389/fmicb.2022.854567
- 20 Zhong X., Guo Z., Yang H., Peng L., Xie Y., Wong T.Y., Lai S.T., Guo Z. Amino terminus of the SARS coronavirus protein 3a elicits strong, potentially protective humoral responses in infected patients. // *J Gen Virol*. – 2006. 87(Pt 2). –P.369-373. <https://doi.org/10.1099/vir.0.81078-0>
- 21 Tamura K., Stecher G., and Kumar S. (2021). MEGA 11: Molecular Evolutionary Genetics Analysis Version 11. // *Molecular Biology and Evolution*. – 2021. 38(7). – P.3022-3027. doi:10.1093/molbev/msab120

- 22 Tamura K., Nei M., Kumar S. Prospects for inferring very large phylogenies by using the neighbor-joining method. // Proc Natl Acad Sci USA. – 2004. 101(30). – P.11030-5. <https://doi.org/10.1073/pnas.0404206101>
- 23 Tzou P.L., Tao K., Nouhin J., Rhee S.Y., Hu B.D., Pai S., Parkin N., Shafer R.W. Coronavirus Antiviral Research Database (CoV-RDB): An Online Database Designed to Facilitate Comparisons between Candidate Anti-Coronavirus Compounds. // Viruses. – 2020. 12(9). – P. 1006. doi:10.3390/v12091006
- 24 Chu D.K.W., Hui K.P.Y., Gu H., Ko R.L.W., Krishnan P., Ng D.Y.M., Liu G.Y.Z., Wan C.K.C., Cheung M.C., Ng K.C., Nicholls J.M., Tsang D.N.C., Peiris M., Chan M.C.W., Poon L.L.M. Introduction of ORF3a-Q57H SARS-CoV-2 Variant Causing Fourth Epidemic Wave of COVID-19, Hong Kong, China. // Emerg Infect Dis. – 2021. 27(5). – P.1492-1495. doi:10.3201/eid2705.210015
- 25 Kern D.M., Sorum B., Mali S.S., Hoel C.M., Sridharan S., Remis J.P., Toso D.B., Kotecha A., Bautista D.M., Brohawn S.G. Cryo-EM structure of SARS-CoV-2 ORF3a in lipid nanodiscs. // Nat Struct Mol Biol. – 2021. 28(8). – P.702. doi:10.1101/2020.06.17.156554
- 26 Desingu P.A., Nagarajan K., Dhama K. Emergence of Omicron third lineage BA.3 and its importance. // J Med Virol. – 2022. 94(5). – P.1808-1810. doi:10.1002/jmv.27601
- 27 Abdullah F., Myers J., Basu D., Tintinger G., Ueckermann V., Mathebula M., Ramlall R., Spoor S., de Villiers T., Van der Walt Z., Cloete J., Soma-Pillay P., Rheeder P., Paruk F., Engelbrecht A., Lalloo V., Myburg M., Kistan J., van Houghenouck-Tulleken W., Boswell M.T., Gray G., Welch R., Blumberg L., Jassat W. Decreased severity of disease during the first global omicron variant covid-19 outbreak in a large hospital in tshwane, south africa. // Int J Infect Dis. – 2022. – P.116:38-42. doi:10.1016/j.ijid.2021.12.357
- 28 Davarinejad H., Huang Y.C., Mermaz B., LeBlanc C., Poulet A., Thomson G., Joly V., Muñoz M., Arvanitis-Vigneault A., Valsakumar D., Villarino G., Ross A., Rotstein B.H., Alarcon E.I., Brunzelle J.S., Voigt P., Dong J., Couture J.F., Jacob Y. The histone H3.1 variant regulates TONSOKU-mediated DNA repair during replication. // Science. – 2022. 375(6586). – P.1281-1286. doi:10.1126/science.abm5320

СЕКВЕНИРОВАНИЕ И ФИЛОГЕНЕТИЧЕСКИЙ АНАЛИЗ ГЕНА ORF3A ВАРИАНТА B.1.1.7 ВИРУСА SARS-COV-2

Б.С. Усербаев *, Н.С. Кожабергенов , А.М. Мелисбек ,
М.Ж. Ширинбеков , А.И. Тагайев , Е.Д. Бурашев 

«Научно-исследовательский институт проблем биологической безопасности», МЗ РК
пгт. Гвардейский

Аннотация: Проблема «новых» инфекционных болезней в настоящее время приобрела особую актуальность для национальных и международных систем здравоохранения. Одной из таких инфекций является коронавирус SARS-CoV-2. В связи с быстрым распространением нового типа коронавируса в мире, важное место занимает проведение молекулярно-генетических исследований данного вируса. Это поможет понять природу вируса и разработать противовирусные препараты с целью профилактики заболевания. Цель - идентификация миссенс-мутации в гене ORF3a исследуемого коронавирусного штамма SARS-CoV-2/human/KAZ/Britain. В данной работе представлены результаты наработки и секвенирования гена ORF3a коронавирусного штамма SARS-CoV-2/human/KAZ/Britain. Для наработки и секвенирования методом Сэнгера гена ORF3a было разработано 4 пары праймеров, с перекрытием 100-150 п.о. Анализированы изменения в гене ORF3a по сравнению с другими штаммами, данные которых получены из базы данных GISAID. В результате полученные последовательности аминокислот гена ORF3a варианта B.1.1.7

были сравнены с эталонным штаммом *hCoV-19/Wuhan/Hu-1/2019*, где была определена одна мутация в позиции 149: W (триптофан) → L (лейцин). Также определена филогенетическая принадлежность исследуемого штамма вируса, где определена генетическая дистанция между исследованными штаммами вируса *SARS-CoV-2*. Таким образом, наибольшее сходство с исследуемым штаммом были:

hCoV-19/Wuhan/Hu-1/2019,
hCoV-19/England/MILK-9E05B3/2020,
hCoV-19/Japan/IC-0446/2020,
hCoV-19/Germany/NI-IOV-MHH15/2020,

а штамм *hCoV-19/Argentina/PAIS-F0418/2021* значительно дистанцировал от штамма *SARS-CoV-2/human/KAZ/Britain*.

Ключевые слова: *SARS-CoV-2*; мутация; ПЦР; праймер; секвенирование

SARS-COV-2 ВИРУСЫ В.1.1.7 ВАРИАНТЫНЫҢ ORF3А ГЕНІН СЕКВЕНИРЛЕУ ЖӘНЕ ФИЛОГЕНЕТИКАЛЫҚ ТАЛДАУ

Б.С. Усербаев , Н.С. Кожабегенов , А.М. Мелисбек ,
 М.Ж. Ширинбеков , А.И. Тагайев , Е.Д. Бурашев 

«Биологиялық қауіпсіздік проблемаларының ғылыми-зерттеу институты» ДМ ҚР
 Гвардейск құк.

Аннотация: «Жаңа» жұқпалы аурулар проблемасы қазіргі уақытта ұлттық және халықаралық денсаулық сақтау саласы үшін ерекше өзектілікке ие. Осындай инфекциялардың бірі-жаңа *SARS-CoV-2* коронавирусы. Коронавирустың жаңа түрінің әлемде тез таралуына байланысты вирус биологиясына молекулалық - генетикалық зерттеулер жүргізу маңызды орын алады. Бұл вирустың табиғатын түсінуге және аурудың алдын алу үшін вирусқа қарсы препараттарды жасауға көмектеседі. Жұмыстың мақсаты - зерттелетін *SARS-CoV-2/human/KAZ/Britain* коронавирустық штамының ORF3а геніндегі миссенс-мутацияны анықтау. Бұл жұмыста *SARS-CoV-2/human/KAZ/Britain* коронавирус штамының ORF3а генінің амплификациясы мен секвенирлеу нәтижелері ұсынылған. ORF3а генін амплификация және Сэнгера әдісімен секвенирлеу үшін қабаттасуы 100 – 150 ж.н. құрайтын 4 жұп праймер жасалынды. GISAID деректер қорынан алынған басқа штамдармен салыстырғанда ORF3а геніндегі өзгерістер талданды. В.1.1.7 нұсқасы ORF3а генінің аминқышқылдарының тізбегін эталондық *hCoV-19/Wuhan/Hu-1/2019* штамымен салыстырылды, нәтижесінде 149 позициясында бір мутация анықталды: W (триптофан) → L (лейцин). Сондай-ақ зерттелетін вирус штамының филогенетикалық сәйкестілігі мен *SARS-CoV-2* вирустарының зерттелген штамдарының арасындағы генетикалық қашықтығы анықталды. Зерттелген штамға ең үлкен ұқсастық:

hCoV-19/Wuhan/Hu-1/2019,
hCoV-19/England/MILK-9E05B3/2020,
hCoV-19/Japan/IC-0446/2020,
hCoV-19/Germany/NI-IOV-MHH15/2020,

ал *hCoV-19/Argentina/PAIS-F0418/2021* штамы *SARS-CoV-2/human/KAZ/Britain* штамынан айтарлықтай алыстады.

Кілтті сөздер: *SARS-CoV-2*; мутация; ПЦР; праймер; секвенирования